

Online Submissions: http://www.wjgnet.com/2150-5330office wjgp@wjgnet.com doi:10.4291/wjgp.v2.i1.1 World J Gastrointest Pathophysiol 2011 February 15; 2(1): 1-14 ISSN 2150-5330 (online) © 2011 Baishideng. All rights reserved.

EDITORIAL

# Therapeutic potential of curcumin in gastrointestinal diseases

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Author contributions: Rajasekaran SA solely contributed to this paper.

Supported by ACS Grant RSG-09-021-01-CNE and The Nemours Foundation

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Telephone: +1-302-6516538 Fax: +1-302-6514827 Received: September 3, 2010 Revised: November 25, 2010 Accepted: December 2, 2010

Published online: February 15, 2011

# Abstract

Curcumin, also known as diferuloylmethane, is derived from the plant Curcuma longa and is the active ingredient of the spice turmeric. The therapeutic activities of curcumin for a wide variety of diseases such as diabetes, allergies, arthritis and other chronic and inflammatory diseases have been known for a long time. More recently, curcumin's therapeutic potential for preventing and treating various cancers is being recognized. As curcumin's therapeutic promise is being explored more systematically in various diseases, it has become clear that, due to its increased bioavailability in the gastrointestinal tract, curcumin may be particularly suited to be developed to treat gastrointestinal diseases. This review summarizes some of the current literature of curcumin's anti-inflammatory, anti-oxidant and anticancer potential in inflammatory bowel diseases, hepatic fibrosis and gastrointestinal cancers.

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Key words: Curcumin; Inflammation; Cancer; Inflamma-

tory bowel disease; Liver fibrosis; Gastrointestinal disease; Apoptosis

**Peer reviewers:** Mária Bagyánszki, PhD, University of Szeged, Department of Physiology, Anatomy and Neuroscience, Közép fasor 52, Szeged, H-6726, Hungary; Jing-Bo Zhao, Associate Professor, Mech-Sense, Research House, AalborgHospital, Adr. Skovvej 15, Aalborg 9000, Denmark

Rajasekaran SA. Therapeutic potential of curcumin in gastrointestinal diseases. *World J Gastrointest Pathophysiol* 2011; 2(1): 1-14 Available from: URL: http://www.wjgnet.com/ 2150-5330/full/v2/i1/1.htm DOI: http://dx.doi.org/10.4291/ wjgp.v2.i1.1

# INTRODUCTION

Curcumin (Curcuma longa) is the active ingredient of the spice turmeric, used in cooking in India and other regions of Asia. It has a long history as an herbal remedy for a variety of diseases and was used in Indian and Chinese traditional medicine as early as 700 AD. Multiple therapeutic activities have been attributed to curcumin mostly because of its anti-inflammatory and anti-oxidant effects. As such, curcumin was predominantly used to treat inflammatory conditions including bronchitis, colds, parasitic worms, leprosy, arthritis and inflammations of bladder, liver, kidney and skin, and to improve symptoms such as fever and diarrhea. In addition, curcumin is thought to have beneficial effects in diseases of the neurological system including Alzheimer's disease<sup>[1]</sup>. Most recently, curcumin has demonstrated chemopreventive and anticancer effects in several human cancers<sup>[2,3]</sup>. In particular, curcumin may slow the growth of gastrointestinal cancers including esophageal, mouth, intestinal, stomach and colon, probably due to its increased bioavailability in the gastrointestinal tract. Curcumin has also been suggested as a remedy for liver and other digestive diseases such as irritable bowel syndrome, colitis, Crohn's disease and bacterial and parasitic diseases. This review seeks to summarize some of the cellular effects of curcumin and explores the possibilities of curcumin in treating gastrointestinal diseases (Figure 1).

# CURCUMIN AND CURCUMIN ANALOGUES

Curcumin, a hydrophobic polyphenol, is the yellow pigment in the Indian spice turmeric derived from the rhizome of the herb Curcuma longa. Curcumin is also known as diferuloylmethane and chemically is a bis- $\alpha$ ,  $\beta$ -unsaturated *B*-diketone. Differing in methoxy substitutions on the aromatic ring, turmeric contains three natural analogues, the so-called curcuminoids, with curcumin being the most abundant (77%) and the less common demethoxycurcumin (17%) and bisdemethoxycurcumin (3%)<sup>[4]</sup>. Curcumin, demethoxycurcumin and bisdemethoxycurcumin differ in their chemical structures only with regard to methoxy substitutions and, while the overall biological activities of these curcuminoids including antioxidant, anti-inflammatory, anti-microbial and anti-carcinogenic properties appear to be comparable, differences in their efficiencies have been observed<sup>[4]</sup>. In addition, several other naturally occurring bioactive compounds have some structural similarity with curcumin such as ferulic acid, cinnamic acid, caffeic acid, capsaicin and gingerol<sup>[4]</sup>. Like curcuminoids, these compounds possess antioxidant, antiinflammatory and anti-tumor activity. However, whether and how the differences in these compounds can be exploited pharmacologically remains to be determined.

In general, natural products are a rich source of potentially therapeutic drugs but many natural products have to be structurally modified and optimized to become useful pharmacological agents. In the case of curcumin, the poor aqueous solubility and relatively low bioavailability have been major obstacles for its clinical development as a therapeutic drug<sup>[5]</sup>. To enhance the solubility and stability and to increase its therapeutic potential, various curcumin analogs have been synthesized. Based on the modification of the basic structure, these compounds are being broadly grouped as "curcumin derivatives" (retain the basic structural features of curcumin), "curcumin analogues" (all other compounds with some structural analogy to curcumin) and "metal complexes of curcumin." In contrast to curcumin analogues, often not obtained from curcumin directly, curcumin derivatives are generally synthesized by derivatization starting from curcumin. Modifications include acetylation, alkylation, glycosylation and amino acetylation of the phenolic hydroxyl group; demethylation of the methoxy groups; or acetylation, alkylation and substitution of the reactive methylene group of the linker (reviewed in<sup>[4]</sup>). Although comparison of the potency of curcumin and its derivatives among different studies is difficult due to the variety of assays and assay systems used, some general trends have emerged regarding the function of the different reactive groups. For example, the phenolic hydroxyl groups are important for curcumin's antioxidant activities, the C7 linker and its carbonyl functions are important for anti-inflammatory activity and the conjugated enones have been shown to act as Michael acceptors for curcumin's anti-cancer activity (for a comprehensive review see<sup>[4,6]</sup>). While the number of curcumin derivatives and analogues is increasing, one of the better known is diphenyl difluoroketone (EF24), a fluorinated analogue that does not have a phenolic hydroxyl group and possesses only one enone. EF24 has been reported to have greater biological activity and bioavailability than curcumin without increased toxicity and it also seems to possess a distinctive mechanism of action from curcumin<sup>[7]</sup>. While EF24 has been shown to have a more potent anti-cancer activity than curcumin, its therapeutic potential in other diseases remains to be determined.

# BIOLOGICAL ACTIVITIES AND MOLECULAR TARGETS OF CURCUMIN

Curcumin has been ascribed a multitude of therapeutic activities and has been associated with suppression of inflammation, angiogenesis, tumorigenesis, and diabetes, and with therapeutic effects in diseases of the cardiovascular, pulmonary and neurological systems and of skin and liver. In general, most of these effects can be attributed to the antioxidant, anti-inflammatory and anticancer activities of curcumin. Curcumin is an effective scavenger of reactive oxygen species and reactive nitrogen species<sup>[8]</sup>. Curcumin's protective function against peroxidative damage of biomembranes, known to be a free-radical-mediated chain reaction, has mainly been attributed to the scavenging of the reactive free radicals involved in peroxidation. These scavenging properties of curcumin have also been considered to be responsible for its protective role against oxidative damage of DNA and proteins, believed to be associated with a variety of chronic diseases such as cancer, atherosclerosis, neurodegenerative diseases and aging<sup>[8]</sup>. In addition to its direct antioxidant activity, curcumin may function indirectly as an antioxidant by inhibiting the activity of inflammatory enzymes or by enhancing the synthesis of glutathione. The anti-inflammatory activity of curcumin seems to be comparable to steroidal drugs and non-steroidal drugs such as indomethacin and phenylbutazone<sup>[8]</sup>. Many studies of curcumin's anti-inflammatory properties point to its ability to suppress prostaglandin synthesis by its effect on cyclooxygenase (COX)<sup>[9]</sup>, a key enzyme responsible for the conversion of arachidonic acid to prostaglandins. Curcumin has also been shown to suppress the production of cytokines such as interferon-y (IFN-y), interleukins and tumor necrosis factor (TNF); to inhibit the inducible nitric oxide synthase (iNOS); and to suppress the activation of NF- $\kappa B^{[9,10]}$ .

These pathways affected by curcumin not only play an important role in inflammation; they have also been associated with the development and progression of cancer. Curcumin has promising chemopreventive and therapeutic potential for various cancers including leukemia,



# Oral cancer and cancer of the submandibular gland

C/EBP alpha, COX-2, CYP1A1, CYP1B1, HSP70, IGFBP-5, MAPK/ ERK, MMP-2/9, NF- $\kappa$ B, p38 MAPK, ROS, uPA

#### Liver fibrosis

α-SMA, CD11b, CTGF, MCP-1, MF-κB, PPARγ, procollagen type I , TGF- $β_1$ , TIMPs, TLR-4

#### **Hepatic cancer**

AP-1, HIF-1, MMP-9, NF-κB

#### Inflammatory bowel diseases, Ulcerative colitis, Crohn's disease

Akt, COX-2, IL-1 $\beta$ , IL-10, iNOS, MAPK/ERK, MyD88, NF- $\kappa$ B, p38 MAPK, PGE2, TLR-4, TRPV-1, VCAM-1



#### Esophageal cancer

NF-κB, PGE2

#### Gastric cancer

AID, caspase-3, cyclin D1, cyclin E, NF- $\kappa$ B, PAK1, P-gp, RTKN

#### Pancreatic cancer

Akt, ATM/Chk1, COX-2, CXCR1, CXCR2, EGFR, HO-1, miR-21, miR-22, miR-200, NF-κB, Notch-1, PGE2, ROS, Sp1, Sp3, Sp4, STAT3, survivin/BIRC5, WT1

#### **Intestinal cancer**

Akt/mTOR, Ca<sup>2+</sup>/calmodulin, COX-2, EGFR, IGF-1R, JNK, IL-8, NF-κB, Wnt/β-catenin

Familial adenomatous polyposis COX-2

**Figure 1 Molecular targets of curcumin in gastrointestinal diseases.** These include AID, activation-induced cytidine deaminase; Akt; AP-1: activated protein-1; α-SMA: alpha smooth muscle actin; caspase-3; ATM/Chk1; Ca<sup>2+</sup>/calmodulin; CD11b; C/EBPalpha: CCAAT/enhancer-binding protein alpha; COX-2: cyclooxygenase-2; CTGF: connective tissue growth factor; CXCR1: chemokine receptor 1; CXCR2: chemokine receptor 2; cyclin D1; cyclin E; CYP1A1: cytochrome P-450 A1; CYP1B1: cytochrome P-450 B1; EGFR: epidermal growth factor receptor; HIF-1: hypoxia-inducible factor-1; HO-1: heme oxygenase 1; HSP70: heat shock protein 70; IGF-1R: insulin-like growth factor 1 receptor; IGFBP-5: insulin-like growth factor binding protein-5; IL-1β/8/10: interleukin-1beta/8/10; iNOS: inducible nitric oxide synthase; JNK: C-jun N-terminal kinase; MAPK/ERK: mitogen-activated protein kinase/extracellular receptor kinase; MCP-1: monocyte chemoattractant protein 1; miR-21, -22, 200: microRNA-21, -22, -200; MMP-2/9: matrix metalloproteinase-2/9; mTOR: mammalian target of rapamycin; MyD88: myeloid differentiation primary response gene (88); NF-κB: nuclear factor-kappa B; Notch-1; p38 MAPK: p38 mitogen-activated protein kinase; PAK1: p21-activated kinase 1; PGE2: prostaglandin E2; P-gp: P-glycoprotein; PPAR<sub>γ</sub>: peroxisome proliferator-activated receptor gamma; procollagen type I ; ROS: reactive oxygen species; RTKN: rhotekin; Sp1, -3, -4; STAT3: signal transducers and activators of transcription 3; surviving/BIRC5; TGF-β<sub>1</sub>: transforming growth factor-beta1; TIMPs: tissue inhibitors of MMPs; TLR-4: toll-like receptor-4; TRPV-1: transient potential vanilloid receptor-1; wP-1: wilms' tumor gene 1.

lymphoma and cancers of the gastrointestinal tract, genitourinary system, breast, ovary, head and neck, lung and skin<sup>[2]</sup>. Aside from avoiding well-established risk factors such as smoking or obesity, the chemopreventive effects of curcumin are one of the most promising approaches to reduce the risk of cancer. Curcumin likely exerts its inhibitory effect on cancer development by several mechanisms such as inhibition of carcinogen activation and stimulation of carcinogen detoxification, prevention of oxidative DNA damage and its capacity to reduce inflammation<sup>[11]</sup>. In several culture and *in vivo* models, curcumin has been shown to modulate enzymes that are involved in the metabolic activation of carcinogens (e.g. inhibition of cytochrome P450) and the detoxification and excretion of such compounds (e.g. induction of glutathione-Stransferase). Of cancer therapeutic interest is the ability of curcumin to induce cell cycle arrest and apoptosis specifically in cancer cells and to inhibit angiogenesis and metastasis. Experimental evidence suggests that curcumin modulates major signaling pathways involved in apoptotic cell death by stimulating pro-apoptotic enzymes such as caspases or by inhibiting cell survival pathways such as Akt, NF- $\kappa$ B, AP-1 and JNK<sup>[12]</sup>. In addition, curcumin exhibits anti-proliferative effects by either modulation of oncogenes and tumor suppressor genes and/or through cell cycle regulatory proteins and checkpoints, thus inducing cell cycle arrest followed by apoptosis. One of the major problems associated with cancer progression to an aggressive disease is angiogenesis, the development of new blood vessels that fuel the rapid growth of tumors. One of the prime factors regulating angiogenesis is vascular endothelial growth factor (VEGF). Curcumin has been shown not only to inhibit VEGF but also other angiogenic factors and could prove effective in preventing the progression of cancer to an aggressive disease. Metastasis is one of the main causes of cancer death with invasive tumors. Interestingly, curcumin treatment significantly inhibits matrix metalloproteinases, a family of zinc-dependent endopeptidases that degrade extracellular matrix, allowing tumors to invade<sup>[13]</sup>. Thus curcumin inhibits a variety of critical steps in the early development of cancer as well as in the progression of cancer to an aggressive and deadly disease and has the potential to develop into a potent cancer therapeutic.

Curcumin affects a multitude of molecular targets that include transcription factors, receptors, kinases, inflam-



matory cytokines and other enzymes (for a comprehensive review see<sup>[14]</sup>), which may partly explain its effectiveness against a variety of diseases. However, it is imperative to distinguish between the primary molecular targets of curcumin and the events caused as downstream effects. For example, curcumin regulates expression and function of COX-2 and 5-LOX, which are important mediators of inflammation by pleiotropic mechanisms on the transcriptional and post-translational level. But, in recent years, curcumin has been found to bind physically to more than 30 proteins; of these, many are directly relevant to the biological activities of curcumin. Interactors of curcumin include albumin; ions such as Zn<sup>2+</sup>, Cu<sup>2+</sup> and Fe<sup>2+</sup>; transcription factors; growth factor receptors; cytokines; genes involved in cell proliferation and apoptosis; and various other enzymes<sup>[15]</sup>. One of the better characterized interactions is the binding of curcumin to tubulin, where it inhibits microtubule assembly. In HeLa and MCF-7 cells, curcumin induces tubulin aggregation and depolymerizes interphase and mitotic microtubules<sup>[16]</sup>. One interesting feature of curcumin is its ability to cross-link proteins. For example, curcumin has been shown to cross-link the cystic fibrosis chloride channel CFTR, thereby activating the channel<sup>[17]</sup>. Because curcumin has such a wide range of target proteins, it remains to be determined whether curcumin binds to a specific structural motif. Identification of curcumin's binding motifs will not only be important to understand curcumin's biological roles but also be a major step in developing more specific and effective curcumin analogs for therapy.

# CURCUMIN AND GASTROINTESTINAL CANCERS

One of the major problems of developing curcumin as a pharmacological therapeutic is its low general bioavailability. However, clinical trials suggest that biologically active levels of curcumin can be achieved in the gastrointestinal tract after oral delivery. This increased availability together with its anti-cancer activity shows promise for curcumin to be developed as a chemotherapeutic agent for gastrointestinal cancers.

#### Oral cavity and submandibular gland

Although studies on curcumin in cancers of the oral cavity are very limited, curcumin has shown some promise in the prevention of oral carcinogenesis. In the 7, 12-dimethylbenz[a]anthracene (DMBA) hamster buccal pouch model of carcinogenesis, curcumin alone<sup>[18,19]</sup> or when administered together with piperine<sup>[20]</sup> significantly reduced the formation of oral carcinoma, probably due to its anti-lipidperoxidative and antioxidant potential together with its effect on modulating carcinogen detoxification. In studies in patients with oral leukoplakia, considered a premalignant lesion, an increase in vitamin C and E suggested that curcumin could mediate anticancer activities by increasing the local and systemic anti-oxidant status, thus preventing lipid peroxidation and DNA damage<sup>[21]</sup>. Other studies suggested that curcumin might act as an oral cavity chemopreventive agent via its ability to inhibit carcinogen activation by inducing expression of and increasing the function of cytochrome P-450 (CYP) 1A1 and/or CYP1B1<sup>[22]</sup>. In leukoplakia cells (MSK-Leuk1s) but not in immortalized oral mucosa epithelial cells (NO M9-CT), curcumin inhibited cell growth by affecting the translation machinery and inhibiting cap-dependent translation<sup>[23]</sup>. In oral epithelial GNM cells, curcumin reduced the expression of heat shock protein 70 (HSP70). High levels of HSP70 protein have been associated with tumor progression<sup>[24]</sup>. Chang *et al*<sup>[25]</sup> reported that in oral keratinocytes, curcumin activates p38, which, in turn, activated the C/EBPalpha (CCAAT/enhancer-binding protein alpha) transactivator to induce insulin-like growth factor binding protein-5 (IGFBP-5). IGFBP-5 upregulation is associated with the suppression of oral cancer cell tumorigenesis in xenografts. In the invasive oral squamous carcinoma cell line YD-10B, curcumin exhibited anti-motility activity which was mediated by the inhibition of MAPK/ERK and NF-KB signaling and consequently down-regulation of proteolytic enzymes such as urokinase-type plasminogen activator (uPA) and matrix metalloproteinases (MMP)-2/9<sup>[26]</sup>. Furthermore, curcumin-abrogated smokeless tobacco induced activation of NF-KB and expression of COX-2 in oral premalignant and cancer cells in vitro<sup>[27]</sup>. Interestingly, in three independent studies in human submandibular gland adenocarcinoma cells (HSG), curcumin induced apoptosis through generation of reactive oxygen species (ROS)<sup>[28-31]</sup>, suggesting that curcumin can induce cell death in these cancer cells.

## Esophagus

Esophageal cancer consists of two types, squamous cell carcinoma and adenocarcinoma. Even with current treatment, overall survival in patients with esophageal cancer remains poor. While there is a desperate need of innovative treatment strategies for esophageal cancer, only a few studies have tested whether curcumin could be a potential candidate. In Flo-1 and OE33 adenocarcinoma cell lines, curcumin inhibited NF-KB activity and induced apoptosis. Curcumin also enhanced 5-fluorouracil (5-FU) and cisplatin (CDDP)-mediated chemosensitivity<sup>[32]</sup>. In a different study, curcumin induced cell death in the two adenocarcinoma cell lines OE33 and OE19 as well as in two squamous cell carcinoma cell lines OE21 and KYSE450 in a dose-dependent manner, probably by inhibiting the ubiquitin-proteasome system  $^{\left[ 33\right] }.$  In the squamous cell carcinoma cell line HKESC-1, curcumin partially reversed the mitogenic effect of prostaglandin E2 (PGE2) that has been implicated in esophageal squamous cell carcinoma growth<sup>[34]</sup>. In addition to its potential chemotherapeutic effect, curcumin might have chemopreventive effects in esophageal cancer. In studies of N-nitrosomethylbenzylamine (NMBA)-induced esophageal carcinogenesis in rats, curcumin inhibited incidence and multiplicity of preneoplastic lesions when given during the post initiation phase and the initiation phase<sup>[35]</sup>.

# Stomach

Helicobacter pylori (H. pylori) infection of gastric epithelial cells is one of the key mechanisms in the development of gastric cancer. One of the proposed molecular mechanisms of *H. pylori*-induced carcinogenesis is the anomalous expression of activation-induced cytidine deaminase (AID), a process that involves NF- $\kappa$ B activation by H. pylon<sup>[36]</sup>. Interestingly, curcumin downregulated H. pyloriinduced AID expression via the inhibition of the NF-KB pathway<sup>[37]</sup>. In addition, curcumin has direct antimicrobial activity against H. pylori and eradicated H. pylori in vitro and in vivo<sup>[38,39]</sup>. Thus, curcumin could be considered as a potential chemopreventive agent against H. pylori-induced gastric carcinogenesis<sup>[37-40]</sup>. Another pathway that has been implicated in gastric tumorigenesis and chemoresistance is one involving Rho, the Rho effector rhotekin (RTKN) and NF- $\!\kappa B^{[41]}\!.$  Curcumin was able to block the RTKNmediated anti-apoptotic effect in AGS cells<sup>[41]</sup>, a cell line that has been previously used to show that curcumin can inhibit the growth of gastric carcinoma cells<sup>[42]</sup>. In cultured gastric cancer cells (BGC823, SGC7901, MKN1 and MGC803), curcumin inhibited invasion and proliferation by downregulating p21-activated kinase 1 (PAK1) activity and cyclin D1 expression<sup>[43]</sup>. Reduced cyclin D and E levels were also found in KATO-III gastric cancer cells upon induction of apoptosis by curcumin<sup>[44]</sup>. In addition, curcumin can reverse the multidrug resistance in the resistant human gastric carcinoma SGC7901/VCR cell line, which might be associated with a decrease in P-glycoprotein (P-gp) function and expression and the promotion of caspase-3 activation<sup>[45]</sup>, suggesting that curcumin might have chemotherapeutic effects. In addition, chemopreventive effects of curcumin have been demonstrated in N-methyl-N'-nitro-N-nitrosoguanidine (MN NG)-induced glandular stomach carcinogenesis<sup>[46-51]</sup> and in benzo[a]pyrene (BaP)-induced forestomach cancer in mice<sup>[52]</sup>. A biological effect of curcumin in chemoprevention of cancer has also been suggested from phase I clinical trials<sup>[53]</sup>.

## Intestine

Of all the gastrointestinal cancers, curcumin's anti-cancer potential has been best characterized in intestinal cancers using cultured tumor cells and in vivo animal models. While Phase I and II clinical trials of curcumin are ongoing and curcumin is well tolerated and pharmacologically active concentrations can be achieved in colorectal tissues in patients after oral administration, conclusive studies as to its anti-cancer activity in patients remain to be done<sup>[54-57]</sup>. Chemopreventive effects of curcumin have been demonstrated in transgenic mouse models and in tumor xenografts. In the 1,2-dimethylhydrazine (DMH)<sup>[58,59]</sup> and the azoxymethane (AOM)-induced<sup>[50,60-64]</sup> rat colon cancer model, curcumin reduced the number of aberrant crypt foci<sup>[59,63]</sup> or inhibited tumor development<sup>[50,58,60-63]</sup>. Treatment of HCT-116 colon tumor-bearing mice with curcumin resulted in reduced tumor growth<sup>[65]</sup>. In some tumor models, curcumin analogs have been suggested to

be more efficient than curcumin; GO-Y030 was found to be more potent than curcumin in inducing apoptosis in cultured human colorectal cancer cells<sup>[66]</sup> and increased the lifespan in Apc(580D+) mice<sup>[66,67]</sup>. EF24 significantly suppressed the tumor growth of HCT-116 colon cancer xenografts<sup>[68]</sup>.

Curcumin may also enhance the effectiveness of current chemotherapeutics and, in combination with chemotherapy, may provide a superior strategy for treatment of gastrointestinal cancers<sup>[ $\delta$ ]</sup>. In APC (Min/+) mice, the combination therapy of curcumin and dasatinib was highly effective in causing regression of intestinal adenomas<sup>[/0]</sup>. The combination of curcumin with resveratrol was found to be more effective in inhibiting the growth of HCT-116 cells in vitro and in vivo in SCID xenografts when compared with resveratrol alone<sup>[71]</sup>. In orthotopic colorectal cancer cell xenografts, curcumin sensitized colorectal cancer to the anti-tumor and anti-metastatic effects of capecitabine by suppressing NF-KB signaling<sup>[72]</sup>. Liposomal curcumin in combination with oxaliplatin significantly inhibited the growth of LoVo and Colo205 xenografts and exhibited anti-angiogenic effects<sup>[73]</sup>. Studies in cultured colon cancer cells found that curcumin can potentiate the pro-apoptotic effects of sulindac sulfone<sup>174]</sup>, 5-fluorouracil<sup>[75,76]</sup> and oxaliplatin<sup>[76,77]</sup>.

In addition to its well-characterized function in suppressing NF- $\kappa$ B and COX-2 signaling and other wellknown mechanisms of inducing apoptosis<sup>[78-82]</sup>, many more molecular targets of curcumin have been identified in colon cancer cell lines. These include inhibition of signaling pathways like EGFR<sup>[76,83-85]</sup>, IGF-1R<sup>[76,85]</sup>, Akt/ mTOR<sup>[86]</sup>, Wnt/ $\beta$ -catenin<sup>[87-89]</sup>, c-jun N-terminal kinase (JNK)<sup>[90]</sup> and Ca<sup>2+</sup>/calmodulin<sup>[91]</sup>. In addition, curcumin may suppress the expression of factors with procarcinogenic effects such as neurotensin-induced IL-8 expression<sup>[92]</sup>. Curcumin has also been shown to inhibit the proteasome activity and induce apoptosis in human colon cancer cells *in vitro* and *in viva*<sup>[65]</sup>.

Radiation therapy is an integral part of the preoperative treatment of rectal cancers. Curcumin has been suggested to be a potential radiosensitizing agent with enhanced anti-angiogenic effects in colorectal cancer radiation therapy<sup>[80]</sup>. Furthermore, curcumin blocked the transient inducible NF- $\kappa$ B signaling in several colon cancer cell lines that provides a prosurvival response to radiation<sup>[81]</sup>. It also sensitized HCT-116 xenografts to gammaradiation<sup>[79]</sup>, suggesting that administration of curcumin could be used to reduce the side-effects usually associated with radiation therapy.

One of the intriguing aspects of curcumin is its chemopreventive function in familial adenomatous polyposis (FAP), which provides an extended opportunity for intervention and cancer prevention. Indeed, combination treatment with curcumin and quercetin appeared to reduce the number and size of ileal and rectal adenomas in patients with FAP<sup>[93]</sup>. In the Apc (Min/+) mouse model of FAP, dietary curcumin showed antioxidant effects and reduced COX-2 expression that has been associated with retarda-



tion in adenoma development<sup>[94,95]</sup>. However, while these initial studies are promising, the chemopreventive effects of curcumin in FAP require further investigation.

## Pancreas

Pancreatic cancer is often diagnosed in an advanced stage and is characterized by rapid disease progression, high invasiveness and resistance to chemotherapy. Recent research suggested that curcumin might have antitumor function in pancreatic cancer alone<sup>[96,97]</sup> or in combination with other drugs such as gemcitabine<sup>[98-102]</sup> or celecoxib<sup>[103]</sup>. Systematic administration of a polymeric nanoparticle-encapsulated curcumin blocked tumor growth and metastasis in subcutaneous and orthotopic Pa03C xenograft models of pancreatic cancer<sup>[96]</sup> and liposomal curcumin decreased tumor growth in cultured pancreatic cancer cells<sup>[104]</sup> and in MiaPaCa-2 subcutaneous xenografts<sup>[105]</sup>. Nanoparticle encapsulation of curcumin into a MePEG/ poly-epsilon caprolactone (PCL) diblock copolymeric micelle enhanced the uptake and cytotoxicity in the pancreatic cancer cell lines MiaPaCa-2 and PANC-1<sup>[106]</sup> and polyethylene glycosylated (PEG) curcumin conjugate was more effective in inducing cell cycle arrest and apoptosis when compared with free curcumin<sup>[107]</sup>. Phase II trials of curcumin in patients with advanced pancreatic cancer revealed that curcumin was well-tolerated and had biological activity in some patients<sup>[108]</sup>.

Many of the molecular pathways that have been implicated in the anticancer properties of curcumin in other gastrointestinal tumors have also been identified in pancreatic cancer, including inhibition of NF-KB signaling<sup>[109-111]</sup>. Inhibition of NF- $\kappa$ B was dependent on downregulation of the transcription factors Sp1, Sp3, and Sp4 and was accompanied by the induction of ROS<sup>[109,112]</sup>. Other targets of curcumin include Akt; COX-2; PGE2; micro RNAs miR-200, miR-21, a signature of tumor aggressiveness, and miR-22<sup>[98,112]</sup>, EGFR<sup>[113]</sup>, Notch-1<sup>[114]</sup>, IL-8 receptors CXCR1 and CXCR2<sup>[115]</sup>, STAT3, survivin/ BIRC5<sup>[116]</sup>, ATM/Chk1<sup>[117]</sup>, heme oxygenase-1 (HO-1)<sup>[118]</sup> and the transcription factor Wilms' tumor gene 1 (WT1)<sup>[119]</sup>. STAT3 activity in pancreatic cancer cell lines was also inhibited by GO-Y030, a potent curcumin analog<sup>[120]</sup>. Other curcumin analogs have shown higher potency than curcumin itself in pancreatic cancer. FLLL11 and FLLL12 were substantially more potent in inhibiting cell viability and inducing apoptosis than curcumin<sup>[121]</sup>. The fluorocurcumin analog of curcumin, CDF, showed a more superior bioavailability in pancreatic tissue when compared with curcumin<sup>[122]</sup>.

# Liver

Few patients with hepatocellular carcinoma are diagnosed early and treatment options are very limited. Surgery is currently the most effective treatment. However, recurrence rates are high and long-term survival is poor. Use of radiation and chemotherapy is limited and identifying novel chemopreventive and chemotherapeutic agents is imperative. Several studies have investigated the anti-

carcinogenic activity of curcumin in hepatic cancer cells in vitro. In HepG2 cells, the curcumin analogue GL63 inhibited growth more effectively than curcumin; this result was associated with the activation of ER stress and apoptosis, an effect not observed with curcumin<sup>[123]</sup>. Curcumin induced G2/M arrest and apoptosis in various hepatoma cell lines such as Huh7, Hep3B, SK-Hep-1 and QGY-7703, but in this study, HepG2 cells were reported to be curcumin-insensitive<sup>[124]</sup>. However, several other studies reported effects of curcumin in HepG2 cells, including induction of apoptosis<sup>[125-127]</sup> through mitochondrial hyperpolarization and mitochondrial DNA damage<sup>[125,126]</sup> or through a p53-dependent pathway<sup>[127]</sup>, and inhibition of hypoxia induced angiogenesis through down-regulation of hypoxia-inducible factor HIF-1<sup>[128]</sup>. In Hepa1-6 cells, knockdown of the extracellular matrix metalloproteinase inducer EMMPRIN (CD147) conferred sensitivity to curcumin<sup>[129]</sup> and curcumin blocked the transactivation of the c-met promoter by activated protein 1 (AP-1)<sup>[130]</sup>. In the highly invasive SK-Hep-1 hepatocellular carcinoma cell, curcumin had significant anti-invasion activity which was associated with inhibition of metalloproteinase MMP-9<sup>[131]</sup>. In hepatic cancer HA22T/VGH cells, curcumin alone or in combination with cisplatin or doxorubicin exerted cell growth inhibitory and apoptotic effects, in part due to changes in NF-kB levels<sup>[132]</sup>.

Curcumin exhibited anticancer properties in several in vivo models of hepatocellular carcinoma. In mice treated with N-bis (2-hydroxypropyl) nitrosamine (DHPN), curcumin significantly inhibited liver adenoma formation and growth<sup>[133]</sup> and in mice with diethylnitrosamine (D EN)-induced hepatocarcinogenesis, curcumin reduced tumor multiplicity and incidence<sup>[134]</sup>. In HepG2 xenografts, tetrahydrocurcumin did not show any cytotoxic activity to HepG2 cells even at the highest doses but had antiangiogenic effects<sup>[135-137]</sup>. In an orthotopic implantation model of hepatocellular carcinoma CBO140C12 cells curcumin suppressed intrahepatic metastasis but did not affect the growth of the implanted tumor itself<sup>[138]</sup>. In N-nitrosodiethylamine (DENA)-initiated and phenobarbital-promoted hepatocarcinogenesis in Wistar rats, curcumin showed beneficial effects against oxidative tissue damage during chemically induced hepatocarcinogenesis<sup>[139-141]</sup>, suggesting that curcumin may exhibit potential as a therapeutic in liver cancer.

# **CURCUMIN AND LIVER FIBROSIS**

Liver fibrosis results from chronic damage to the liver due to excessive accumulation of extracellular matrix proteins and is considered a wound healing response to chronic liver injury. The main causes of liver fibrosis are considered to be chronic hepatitis C infection, alcohol abuse and nonalcoholic steatohepatitis. The major collagen-producing cells during chronic liver injury are hepatic stellate cells (HSCs) but the cells also include portal myofibroblasts and myofibroblasts of bone marrow origin and are activated by fibrogenic cytokines such as TG



F- $\beta_1$ , angiotensin II and leptin. In recent years, it has been shown that advanced liver fibrosis can be reversed, thus prompting the development of antifibrotic drugs. It has been shown that curcumin can protect HSCs against activation by leptin and leptin-induced oxidative stress<sup>[142,143]</sup> by interrupting insulin signaling<sup>[144]</sup>, by attenuation of oxidative stress<sup>[144,145]</sup>, by inhibition of LDL-induced activation<sup>[146-148]</sup>, by inhibiting TGF- $\beta_1$  induced  $\alpha$ -smooth muscle actin ( $\alpha$ -SMA) expression and collagen deposition<sup>[149]</sup> and by activation of PPARgamma (peroxisome proliferator-activated receptor gamma)<sup>[150,151]</sup>. At higher concentrations, curcumin induced apoptosis in HSCs<sup>[149,152]</sup>. Increased expression of connective tissue growth factor (CTGF) in HSCs increases ECM production and is mediated by NF- $\kappa$ B. In these cells, curcumin suppressed gene expression of Toll-like receptor-4, thus inhibiting NF-κB and suppressing CTGF expression<sup>[153]</sup>.

In liver fluke-infected hamsters that develop advanced periductal fibrosis, long-term treatment with curcumin reduced fibrosis via inhibition of TIMPs (tissue inhibitors of MMPs)<sup>[154]</sup>. After curcumin feeding, liver damage, cholestasis and fibrosis were reduced in Mdr2 (-/-) mice, a murine model of chronic cholangiopathy, and cholangiocyte proliferation and expression of the activation marker vascular cell adhesion molecule-1 was inhibited<sup>[155]</sup>. In carbon tetrachloride (CCl4)-induced liver damage in Sprague-Dawley rats, curcumin inhibited CCl4-induced NF-KB expression and proinflammatory cytokines and significantly reduced hepatic collagen deposition<sup>[156]</sup>, improved hepatic antioxidant activity<sup>[157,158]</sup> and induced apoptosis in HSCs<sup>[159]</sup>. When liver cirrhosis was induced with thioacetamide, curcumin reduced oxidative stress, inhibited HSC activation and collagen alpha 1 expression and inhibited the development of liver cirrhosis<sup>[160]</sup>. Mice fed with a diet deficient in methionine and choline (MCD) develop nonalcoholic steatohepatitis. In MCD mice, curcumin significantly reduced fibrosis and decreased the intrahepatic expression of monocyte chemoattractant protein1, CD11b, procollagen type I, TIMP-1 and  $\alpha$ -SMA<sup>[161]</sup>. These studies suggest that curcumin might be beneficial in liver fibrosis. However, conclusive clinical trials have still not been reported.

# CURCUMIN AND INFLAMMATORY BOWEL DISEASE

Inflammatory bowel disease (IBD) refers to two chronic inflammatory diseases: ulcerative colitis, an inflammatory disease of the colon mucosa, and Crohn's disease with inflammation that extends much deeper mainly into the ileum. IBD mostly affects young people ages 15 to 30 but it can affect younger children and older adults. Treatment for IBD consists mainly of anti-inflammatory drugs and immunosuppressive agents. Thus, curcumin with its antiinflammatory properties is an ideal drug candidate to possibly treat IBD. In human IBD, one study has achieved encouraging results. In a randomized, multicenter, doubleblind, placebo-controlled trial to assess the efficacy of curcumin as maintenance therapy in patients with quiescent ulcerative colitis, recurrence rates in curcumin-treated patients were significantly lower than in the placebo group<sup>[162]</sup>. In a pilot study when curcumin was administered in an open-label study to five patients with ulcerative proctitis and five patients with Crohn's disease, all patients with proctitis improved and four patients with Crohn's disease had a lowered CDAI score<sup>[163]</sup>. In colonic mucosal biopsies and colonic myofibroblasts isolated from children and adults with active IBD, curcumin reduced p38 MAPK activation, enhanced IL-10 and reduced IL1beta<sup>[164]</sup>.

In recent years, the efficacy of curcumin has been investigated in several experimental models of IBD and has shown encouraging results. However, in IL-10-deficient mice, curcumin demonstrated only limited effectiveness on Th-1-mediated colitis and only moderately improved colonic morphology, suggesting that curcumin' s protective effects are IL-10-dependent<sup>[165]</sup>. A different study suggested that curcumin had potent anti-inflammatory effects in IL-10 deficient mice<sup>[166]</sup> only when it was emulsified in carboxymethyl cellulose, suggesting that bioavailability might be an issue. In multidrug resistance gene deficient (mdr1a-/-) mice that spontaneously develop intestinal inflammation predominantly in the colon, addition of curcumin to the diet significantly reduced the histological signs of colonic inflammation<sup>[167]</sup>. Using mice with dinitrobenzene sulfonic acid (DNBS)-induced colitis, curcumin was shown to attenuate colonic inflammation through a mechanism that correlated with the inhibition of the activation of NF-KB and reduction in p38 MAPK activity<sup>[168]</sup>. In the DNBS mouse model of IBD, curcumin was shown to interact with transient potential vanilloid receptor-1 (TRPV-1) in inflamed tissue to mediate a protective action<sup>[169]</sup>. TRPV-1 has been shown to play an important role in visceral hypersensitivity.

In rats with experimentally induced colitis by intra-rectal administration of trinitrobenzenesulfonic acid (TNBS), curcumin attenuated inflammation through inhibition of toll-like receptor (TLR)-4 and MyD88, a pathway that induces NF- $\kappa$ B leading to inflammation<sup>[170,171]</sup>, and NF-KB activation in colonic mucosa was suppressed<sup>[172]</sup>. Curcumin also inhibited p38 MAPK signaling, downregulated COX-2 and iNOS expression, and increased PGE2 expression in TNBS rats<sup>[173,174]</sup>. However, in TNBSinduced colitis in two different strains of mice, BALB/c and SJL/J, curcumin significantly increased survival, prevented weight loss and normalized disease activity in BA LB/c mice but not SJL/J mice, suggesting that the therapeutic value of curcumin may differ depending on the nature of immune dysregulation in IBD<sup>[175]</sup>. When rats with dinitrochlorobenzene (DNCB)-induced colitis were treated with curcumin, the mice showed a dose-dependent protection against colonic myeloperoxidase (MPO), lipid peroxidation (LPO) and alkaline phosphatase (ALP) activities. At the highest dose, curcumin inhibited DNCBinduced overexpression of NF-KB and iNOS in the colon and was almost equipotent with sulfasalazine in ameliorating colonic damage<sup>[176]</sup>. In the dextran sulfate sodium (DSS)-induced ulcerative colitis rat model, administration of curcumin ahead of the DSS challenge mitigated the injurious effects of DSS<sup>[177]</sup>. A hydroxypropyl-beta cyclodextrin complex of curcumin<sup>[178]</sup> as well as solid lipid microparticles of curcumin<sup>[179]</sup> significantly attenuated the degree of colitis caused by DSS.

Curcumin may not act only on colonic epithelial cells but may also modulate the immune response by inhibiting neutrophil chemotaxis and chemokines<sup>[180]</sup>. Neutrophils are the first cells recruited at the site of inflammation and play a key role in the innate immune responses. In IBD, transepithelial neutrophil migration leads to impaired epithelial barrier function, perpetuation of inflammation and tissue destruction<sup>[180]</sup>. In addition, curcumin may prevent endothelial activation in IBD, critical for binding and recruitment of circulating leukocytes during the inflammatory response. In primary cultures of human intestinal microvascular endothelial cells (HIMEC), curcumin inhibited Akt/MAPK/NF-KB activity and prevented nuclear translocation of the p65 NF- $\kappa B$  subunit following tumor necrosis factor (TNF)- $\alpha$ / lipopolysaccharide (LPS) treatment<sup>[181]</sup>. Curcumin furthermore inhibited the expression of VCAM-1 (vascular cell adhesion molecule-1) in HIMEC cells, which is critical for binding and recruitment of circulating leukocytes<sup>[181]</sup> and reduction of COX-2 and PGE2 and inhibition of MAPK signaling<sup>[78]</sup>. Consistent with these studies in cultured cells, in TNBS-induced colitis in mice, curcumin treatment resulted in a significant reduction of neutrophil invasion<sup>[182]</sup> and CD4 (+) T-cell infiltration<sup>[183]</sup>. Thus, curcumin shows promise as a potential therapeutic agent for the treatment of patients with IBD.

# **BIOAVAILABILITY OF CURCUMIN**

Although the pharmacological safety of curcumin promises a great potential for treatment and prevention of various diseases, the relatively low bioavailability of curcumin is a major hurdle for clinical development. Curtailing curcumin's bioavailability primarily are its poor absorption and rapid metabolism resulting in low serum levels, limited tissue distribution and short half-life. Distribution studies for curcumin showed its preferential accumulation in the intestine, colon and liver and might be one of the major reasons why its most promising in vivo effects have been in gastrointestinal diseases when compared with other organ systems. The liver, and to a lesser extent the intestinal mucosa, were found to be the major organs responsible for metabolism of curcumin<sup>[5]</sup>. Once absorbed, curcumin is readily conjugated to form curcumin glucuronides and curcumin sulfates or it is reduced to hexahydrocurcumin. To date, there is no clear understanding whether these curcumin metabolites are pharmacologically as active as curcumin or whether these conjugates have effects that differ from those of curcumin.

Novel approaches to overcome the problem of low bioavailability of curcumin are being developed. These

approaches include adjuvants that can block the metabolic pathways of curcumin and improved formulations for curcumin delivery such as liposomes, micelles, phospholipid complexes and nanoparticles. Piperine is a known inhibitor of hepatic and intestinal glucuronidation and combined administration of piperine and curcumin greatly enhances curcumin's bioavailability<sup>[184]</sup>. Another agent showing synergistic effects with curcumin is quercetin, used in a clinical study in patients with FAP in which the number of polyps decreased<sup>[93]</sup>.

Encapsulation of curcumin in liposomes with their hydrophilic and hydrophobic properties should offer an excellent drug delivery system. Liposomal curcumin has shown anti-tumor activity in pancreatic carcinoma cells and inhibits pancreatic carcinoma growth<sup>[104]</sup>. *In vivo* studies on colorectal tumor xenografts showed that liposomal curcumin enhanced the antitumor effect of oxaliplatin<sup>[73]</sup>. While these studies showed that liposomal curcumin is bioactive, the increased bioavailability of encapsulated curcumin *in vivo* still needs to be conclusively demonstrated.

Nanoparticle-based systems are one of the most promising delivery systems for curcumin but are still in their infancy. A recent study using a polymer-based nanoparticle of curcumin found it to have a molecular activity similar to free curcumin in pancreatic cell lines<sup>[185]</sup>. Encapsulation of curcumin in a pluronic block copolymer demonstrated a slow and sustained release of curcumin and showed comparable anticancer activity with free curcumin<sup>[186]</sup>. Thus, nanotechnology-based approaches to increase curcumin delivery *in vivo* are evolving and are very promising in overcoming the problem of bioavailability of curcumin, a drug with high potential as a therapeutic for gastrointestinal cancers and inflammatory diseases as well as liver fibrosis.

# **CONCLUSIONS AND PERSPECTIVES**

Curcumin has been used for many centuries to treat various diseases. With its vast array of molecular targets it has shown great potential as a therapeutic agent for various cancer types and for inflammatory conditions. Because of its increased bioavailability in the gastrointestinal tract, gastrointestinal diseases including inflammatory bowel diseases, hepatic fibrosis and gastrointestinal cancers are one of the most investigated diseases demonstrating the therapeutic potential of curcumin. Over many millennia, natural products have often been the only means to treat diseases and injuries. With the advances in molecular biology and combinatorial chemistry during the past few decades, natural products have taken a secondary role in drug discovery. However, in the past few years, the interest in natural drugs as compounds for drug development has been renewed. As a natural product, curcumin has taken precedence over many other compounds and biomedical research on curcumin is everincreasing. As the problems of curcumin absorption, biodistribution, metabolism and elimination are being overcome to enhance its bioavailability, diseases of the gastrointestinal tract will be at the forefront as one of the most promising targets for curcumin therapy.

# ACKNOWLEDGEMENTS

I thank Ms. Sridevi Mony for help with the illustration and Drs. Rajasekaran AK and Seung Joon Lee for critical reading of the manuscript.

# REFERENCES

- Kim J, Lee HJ, Lee KW. Naturally occurring phytochemicals for the prevention of Alzheimer's disease. J Neurochem 2010; 112: 1415-1430
- 2 Anand P, Sundaram C, Jhurani S, Kunnumakkara AB, Aggarwal BB. Curcumin and cancer: an "old-age" disease with an "age-old" solution. *Cancer Lett* 2008; **267**: 133-164
- 3 Goel A, Kunnumakkara AB, Aggarwal BB. Curcumin as "Curecumin": from kitchen to clinic. *Biochem Pharmacol* 2008; 75: 787-809
- 4 Anand P, Thomas SG, Kunnumakkara AB, Sundaram C, Harikumar KB, Sung B, Tharakan ST, Misra K, Priyadarsini IK, Rajasekharan KN, Aggarwal BB. Biological activities of curcumin and its analogues (Congeners) made by man and Mother Nature. *Biochemical Pharmacology* 2008; **76**: 1590-1611
- 5 Anand P, Kunnumakkara AB, Newman RA, Aggarwal BB. Bioavailability of curcumin: problems and promises. *Mol Pharm* 2007; 4: 807-818
- 6 **Mosley CA**, Liotta DC, Snyder JP. Highly active anticancer curcumin analogues. *Adv Exp Med Biol* 2007; **595**: 77-103
- 7 Thomas SL, Zhong D, Zhou W, Malik S, Liotta D, Snyder JP, Hamel E, Giannakakou P. EF24, a novel curcumin analog, disrupts the microtubule cytoskeleton and inhibits HIF-1. *Cell Cycle* 2008; 7: 2409-2417
- 8 Menon VP, Sudheer AR. Antioxidant and anti-inflammatory properties of curcumin. Adv Exp Med Biol 2007; 595: 105-125
- 9 Rao CV. Regulation of COX and LOX by curcumin. Adv Exp Med Biol 2007; 595: 213-226
- 10 **Kim YS**, Young MR, Bobe G, Colburn NH, Milner JA. Bioactive food components, inflammatory targets, and cancer prevention. *Cancer Prev Res (Phila)* 2009; **2**: 200-208
- 11 Surh YJ, Chun KS. Cancer chemopreventive effects of curcumin. Adv Exp Med Biol 2007; 595: 149-172
- 12 **Thangapazham RL**, Sharma A, Maheshwari RK. Multiple molecular targets in cancer chemoprevention by curcumin. *AAPS J* 2006; **8**: E443-E449
- 13 Lin SS, Lai KC, Hsu SC, Yang JS, Kuo CL, Lin JP, Ma YS, Wu CC, Chung JG. Curcumin inhibits the migration and invasion of human A549 lung cancer cells through the inhibition of matrix metalloproteinase-2 and -9 and Vascular Endothelial Growth Factor (VEGF). *Cancer Lett* 2009; 285: 127-133
- Aggarwal BB, Sundaram C, Malani N, Ichikawa H. Curcumin: the Indian solid gold. *Adv Exp Med Biol* 2007; 595: 1-75
- 15 Kunnumakkara AB, Anand P, Aggarwal BB. Curcumin inhibits proliferation, invasion, angiogenesis and metastasis of different cancers through interaction with multiple cell signaling proteins. *Cancer Lett* 2008; 269: 199-225
- 16 Gupta KK, Bharne SS, Rathinasamy K, Naik NR, Panda D. Dietary antioxidant curcumin inhibits microtubule assembly through tubulin binding. *FEBS J* 2006; 273: 5320-5332
- 17 Bernard K, Wang W, Narlawar R, Schmidt B, Kirk KL. Curcumin cross-links cystic fibrosis transmembrane conductance regulator (CFTR) polypeptides and potentiates CFTR channel activity by distinct mechanisms. *J Biol Chem* 2009; 284: 307 54-30765
- 18 Krishnaswamy K, Goud VK, Sesikeran B, Mukundan MA,

Krishna TP. Retardation of experimental tumorigenesis and reduction in DNA adducts by turmeric and curcumin. *Nutr Cancer* 1998; **30**: 163-166

- 19 Li N, Chen X, Liao J, Yang G, Wang S, Josephson Y, Han C, Chen J, Huang MT, Yang CS. Inhibition of 7,12-dimeth ylbenz[a]anthracene (DMBA)-induced oral carcinogenesis in hamsters by tea and curcumin. *Carcinogenesis* 2002; 23: 1307-1313
- 20 Manoharan S, Balakrishnan S, Menon VP, Alias LM, Reena AR. Chemopreventive efficacy of curcumin and piperine during 7,12-dimethylbenz[a]anthracene-induced hamster buccal pouch carcinogenesis. *Singapore Med J* 2009; 50: 139-146
- 21 Rai B, Kaur J, Jacobs R, Singh J. Possible action mechanism for curcumin in pre-cancerous lesions based on serum and salivary markers of oxidative stress. J Oral Sci 2010; 52: 251-256
- 22 **Rinaldi AL**, Morse MA, Fields HW, Rothas DA, Pei P, Rodrigo KA, Renner RJ, Mallery SR. Curcumin activates the aryl hydrocarbon receptor yet significantly inhibits (-)-benzo(a)pyrene-7R-trans-7,8-dihydrodiol bioactivation in oral squamous cell carcinoma cells and oral mucosa. *Cancer Res* 2002; **62**: 5451-5456
- 23 Chakravarti N, Kadara H, Yoon DJ, Shay JW, Myers JN, Lotan D, Sonenberg N, Lotan R. Differential inhibition of protein translation machinery by curcumin in normal, immortalized, and malignant oral epithelial cells. *Cancer Prev Res* (*Phila*) 2010; **3**: 331-338
- 24 Lee SS, Tsai CH, Ho YC, Chang YC. The upregulation of heat shock protein 70 expression in areca quid chewing-associated oral squamous cell carcinomas. *Oral Oncol* 2008; 44: 884-890
- 25 Chang KW, Hung PS, Lin IY, Hou CP, Chen LK, Tsai YM, Lin SC. Curcumin upregulates insulin-like growth factor binding protein-5 (IGFBP-5) and C/EBPalpha during oral cancer suppression. *Int J Cancer* 2010; **127**: 9-20
- 26 Shin HK, Kim J, Lee EJ, Kim SH. Inhibitory effect of curcumin on motility of human oral squamous carcinoma YD-10B cells via suppression of ERK and NF-kappaB activations. *Phytother Res* 2010; 24: 577-582
- 27 Sharma C, Kaur J, Shishodia S, Aggarwal BB, Ralhan R. Curcumin down regulates smokeless tobacco-induced NF-kappaB activation and COX-2 expression in human oral premalignant and cancer cells. *Toxicology* 2006; 228: 1-15
- 28 Atsumi T, Fujisawa S, Tonosaki K. Relationship between intracellular ROS production and membrane mobility in curcumin- and tetrahydrocurcumin-treated human gingival fibroblasts and human submandibular gland carcinoma cells. *Oral Dis* 2005; 11: 236-242
- 29 Atsumi T, Murakami Y, Shibuya K, Tonosaki K, Fujisawa S. Induction of cytotoxicity and apoptosis and inhibition of cyclooxygenase-2 gene expression, by curcumin and its analog, alpha-diisoeugenol. *Anticancer Res* 2005; 25: 4029-4036
- 30 Atsumi T, Tonosaki K, Fujisawa S. Induction of early apoptosis and ROS-generation activity in human gingival fibroblasts (HGF) and human submandibular gland carcinoma (HSG) cells treated with curcumin. *Arch Oral Biol* 2006; 51: 913-921
- 31 Fujisawa S, Atsumi T, Ishihara M, Kadoma Y. Cytotoxicity, ROS-generation activity and radical-scavenging activity of curcumin and related compounds. *Anticancer Res* 2004; 24: 563-569
- 32 Hartojo W, Silvers AL, Thomas DG, Seder CW, Lin L, Rao H, Wang Z, Greenson JK, Giordano TJ, Orringer MB, Rehemtulla A, Bhojani MS, Beer DG, Chang AC. Curcumin promotes apoptosis, increases chemosensitivity, and inhibits nuclear factor kappaB in esophageal adenocarcinoma. *Transl Oncol* 2010; **3**: 99-108
- 33 O'Sullivan-Coyne G, O'Sullivan GC, O'Donovan TR, Piwocka K, McKenna SL. Curcumin induces apoptosis-independent death in oesophageal cancer cells. Br J Cancer 2009; 101: 1585-1595
- 34 Yu L, Wu WK, Li ZJ, Wong HP, Tai EK, Li HT, Wu YC, Cho



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CH. E series of prostaglandin receptor 2-mediated activation of extracellular signal-regulated kinase/activator protein-1 signaling is required for the mitogenic action of prostaglandin E2 in esophageal squamous-cell carcinoma. *J Pharmacol Exp Ther* 2008; **327**: 258-267

- 35 **Ushida J**, Sugie S, Kawabata K, Pham QV, Tanaka T, Fujii K, Takeuchi H, Ito Y, Mori H. Chemopreventive effect of curcumin on N-nitrosomethylbenzylamine-induced esophageal carcinogenesis in rats. *Jpn J Cancer Res* 2000; **91**: 893-898
- 36 Matsumoto Y, Marusawa H, Kinoshita K, Endo Y, Kou T, Morisawa T, Azuma T, Okazaki I-M, Honjo T, Chiba T. Helicobacter pylori infection triggers aberrant expression of activation-induced cytidine deaminase in gastric epithelium. *Nat Med* 2007; 13: 470-476
- 37 Zaidi SF, Yamamoto T, Refaat A, Ahmed K, Sakurai H, Saiki I, Kondo T, Usmanghani K, Kadowaki M, Sugiyama T. Modulation of activation-induced cytidine deaminase by curcumin in Helicobacter pylori-infected gastric epithelial cells. *Helicobacter* 2009; 14: 588-595
- 38 De R, Kundu P, Swarnakar S, Ramamurthy T, Chowdhury A, Nair GB, Mukhopadhyay AK. Antimicrobial activity of curcumin against Helicobacter pylori isolates from India and during infections in mice. *Antimicrob Agents Chemother* 2009; 53: 1592-1597
- 39 Mahady GB, Pendland SL, Yun G, Lu ZZ. Turmeric (Curcuma longa) and curcumin inhibit the growth of Helicobacter pylori, a group 1 carcinogen. *Anticancer Res* 2002; 22: 4179-4181
- 40 Munzenmaier A, Lange C, Glocker E, Covacci A, Moran A, Bereswill S, Baeuerle PA, Kist M, Pahl HL. A secreted/shed product of Helicobacter pylori activates transcription factor nuclear factor-kappa B. J Immunol 1997; 159: 6140-6147
- 41 Liu CA, Wang MJ, Chi CW, Wu CW, Chen JY. Rho/Rhotekin-mediated NF-kappaB activation confers resistance to apoptosis. *Oncogene* 2004; 23: 8731-8742
- 42 **Koo JY**, Kim HJ, Jung KO, Park KY. Curcumin inhibits the growth of AGS human gastric carcinoma cells in vitro and shows synergism with 5-fluorouracil. *J Med Food* 2004; 7: 117-121
- 43 Cai XZ, Wang J, Li XD, Wang GL, Liu FN, Cheng MS, Li F. Curcumin suppresses proliferation and invasion in human gastric cancer cells by downregulation of PAK1 activity and cyclin D1 expression. *Cancer Biol Ther* 2009; 8: 1360-1368
- 44 **Moragoda L**, Jaszewski R, Majumdar AP. Curcumin induced modulation of cell cycle and apoptosis in gastric and colon cancer cells. *Anticancer Res* 2001; **21**: 873-878
- 45 Tang XQ, Bi H, Feng JQ, Cao JG. Effect of curcumin on multidrug resistance in resistant human gastric carcinoma cell line SGC7901/VCR. *Acta Pharmacol Sin* 2005; 26: 1009-1016
- 46 Azuine MA, Bhide SV. Chemopreventive effect of turmeric against stomach and skin tumors induced by chemical carcinogens in Swiss mice. *Nutr Cancer* 1992; 17: 77-83
- 47 Azuine MA, Bhide SV. Adjuvant chemoprevention of experimental cancer: catechin and dietary turmeric in forestomach and oral cancer models. *J Ethnopharmacol* 1994; 44: 211-217
- 48 Azuine MA, Kayal JJ, Bhide SV. Protective role of aqueous turmeric extract against mutagenicity of direct-acting carcinogens as well as benzo [alpha] pyrene-induced genotoxicity and carcinogenicity. J Cancer Res Clin Oncol 1992; 118: 447-452
- 49 Deshpande SS, Ingle AD, Maru GB. Inhibitory effects of curcumin-free aqueous turmeric extract on benzo[a]pyrene-induced forestomach papillomas in mice. *Cancer Lett* 1997; 118: 79-85
- 50 Huang MT, Lou YR, Ma W, Newmark HL, Reuhl KR, Conney AH. Inhibitory effects of dietary curcumin on forestomach, duodenal, and colon carcinogenesis in mice. *Cancer Res* 1994; 54: 5841-5847
- 51 **Ikezaki S**, Nishikawa A, Furukawa F, Kudo K, Nakamura H, Tamura K, Mori H. Chemopreventive effects of curcumin on glandular stomach carcinogenesis induced by N-methyl-N'

-nitro-N-nitrosoguanidine and sodium chloride in rats. Anticancer Res 2001; 21: 3407-3411

- 52 Singh SV, Hu X, Srivastava SK, Singh M, Xia H, Orchard JL, Zaren HA. Mechanism of inhibition of benzo[a]pyreneinduced forestomach cancer in mice by dietary curcumin. *Carcinogenesis* 1998; 19: 1357-1360
- 53 Cheng AL, Hsu CH, Lin JK, Hsu MM, Ho YF, Shen TS, Ko JY, Lin JT, Lin BR, Ming-Shiang W, Yu HS, Jee SH, Chen GS, Chen TM, Chen CA, Lai MK, Pu YS, Pan MH, Wang YJ, Tsai CC, Hsieh CY. Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Res* 2001; **21**: 2895-2900
- 54 Hsu CH, Cheng AL. Clinical studies with curcumin. *Adv Exp Med Biol* 2007; 595: 471-480
- 55 Johnson JJ, Mukhtar H. Curcumin for chemoprevention of colon cancer. *Cancer Lett* 2007; **255**: 170-181
- 56 Sharma RA, McLelland HR, Hill KA, Ireson CR, Euden SA, Manson MM, Pirmohamed M, Marnett LJ, Gescher AJ, Steward WP. Pharmacodynamic and pharmacokinetic study of oral Curcuma extract in patients with colorectal cancer. *Clin Cancer Res* 2001; 7: 1894-1900
- 57 Sharma RA, Steward WP, Gescher AJ. Pharmacokinetics and pharmacodynamics of curcumin. Adv Exp Med Biol 2007; 595: 453-470
- 58 Devasena T, Rajasekaran KN, Gunasekaran G, Viswanathan P, Menon VP. Anticarcinogenic effect of bis-1,7-(2-hydro-xyphenyl)-hepta-1,6-diene-3,5-dione a curcumin analog on DMH-induced colon cancer model. *Pharmacol Res* 2003; 47: 133-140
- 59 Shpitz B, Giladi N, Sagiv E, Lev-Ari S, Liberman E, Kazanov D, Arber N. Celecoxib and curcumin additively inhibit the growth of colorectal cancer in a rat model. *Digestion* 2006; 74: 140-144
- 60 Kawamori T, Lubet R, Steele VE, Kelloff GJ, Kaskey RB, Rao CV, Reddy BS. Chemopreventive effect of curcumin, a naturally occurring anti-inflammatory agent, during the promotion/progression stages of colon cancer. *Cancer Res* 1999; 59: 597-601
- 61 Pereira MA, Grubbs CJ, Barnes LH, Li H, Olson GR, Eto I, Juliana M, Whitaker LM, Kelloff GJ, Steele VE, Lubet RA. Effects of the phytochemicals, curcumin and quercetin, upon azoxymethane-induced colon cancer and 7,12-dimethylbenz[a]anthracene-induced mammary cancer in rats. *Carcinogenesis* 1996; 17: 1305-1311
- 62 **Rao CV**, Rivenson A, Simi B, Reddy BS. Chemoprevention of colon carcinogenesis by dietary curcumin, a naturally occurring plant phenolic compound. *Cancer Res* 1995; **55**: 259-266
- 63 Volate SR, Davenport DM, Muga SJ, Wargovich MJ. Modulation of aberrant crypt foci and apoptosis by dietary herbal supplements (quercetin, curcumin, silymarin, ginseng and rutin). *Carcinogenesis* 2005; 26: 1450-1456
- 64 Wijnands MV, van Erk MJ, Doornbos RP, Krul CA, Woutersen RA. Do aberrant crypt foci have predictive value for the occurrence of colorectal tumours? Potential of gene expression profiling in tumours. *Food Chem Toxicol* 2004; **42**: 1629-1639
- 65 **Milacic V**, Banerjee S, Landis-Piwowar KR, Sarkar FH, Majumdar AP, Dou QP. Curcumin inhibits the proteasome activity in human colon cancer cells in vitro and in vivo. *Cancer Res* 2008; **68**: 7283-7292
- 66 **Cen L**, Hutzen B, Ball S, DeAngelis S, Chen CL, Fuchs JR, Li C, Li PK, Lin J. New structural analogues of curcumin exhibit potent growth suppressive activity in human colorectal carcinoma cells. *BMC Cancer* 2009; **9**: 99
- 67 Shibata H, Yamakoshi H, Sato A, Ohori H, Kakudo Y, Kudo C, Takahashi Y, Watanabe M, Takano H, Ishioka C, Noda T, Iwabuchi Y. Newly synthesized curcumin analog has improved potential to prevent colorectal carcinogenesis in vivo. *Cancer Sci* 2009; **100**: 956-960
- 68 Subramaniam D, May R, Sureban SM, Lee KB, George R,



Kuppusamy P, Ramanujam RP, Hideg K, Dieckgraefe BK, Houchen CW, Anant S. Diphenyl difluoroketone: a curcumin derivative with potent in vivo anticancer activity. *Cancer Res* 2008; **68**: 1962-1969

- 69 Patel BB, Majumdar AP. Synergistic role of curcumin with current therapeutics in colorectal cancer: minireview. *Nutr Cancer* 2009; 61: 842-846
- 70 Nautiyal J, Banerjee S, Kanwar SS, Yu Y, Patel BB, Sarkar FH, Majumdar AP. Curcumin enhances dasatinib-induced inhibition of growth and transformation of colon cancer cells. Int J Cancer 2010; Epub ahead of print
- 71 **Majumdar AP**, Banerjee S, Nautiyal J, Patel BB, Patel V, Du J, Yu Y, Elliott AA, Levi E, Sarkar FH. Curcumin synergizes with resveratrol to inhibit colon cancer. *Nutr Cancer* 2009; **61**: 544-553
- 72 Kunnumakkara AB, Diagaradjane P, Anand P, Harikumar KB, Deorukhkar A, Gelovani J, Guha S, Krishnan S, Aggarwal BB. Curcumin sensitizes human colorectal cancer to capecitabine by modulation of cyclin D1, COX-2, MMP-9, VEGF and CXCR4 expression in an orthotopic mouse model. *Int J Cancer* 2009; **125**: 2187-2197
- 73 Li L, Ahmed B, Mehta K, Kurzrock R. Liposomal curcumin with and without oxaliplatin: effects on cell growth, apoptosis, and angiogenesis in colorectal cancer. *Mol Cancer Ther* 2007; 6: 1276-1282
- 74 Giladi N, Kazanov D, Shpitz B, Aroch I, Kraus S, Arber N. Curcumin potentiates the pro-apoptotic effects of sulindac sulfone in colorectal cancer. *Expert Opin Investig Drugs* 2010; 19 Suppl 1: S117-S124
- 75 Du B, Jiang L, Xia Q, Zhong L. Synergistic inhibitory effects of curcumin and 5-fluorouracil on the growth of the human colon cancer cell line HT-29. *Chemotherapy* 2006; **52**: 23-28
- 76 Patel BB, Sengupta R, Qazi S, Vachhani H, Yu Y, Rishi AK, Majumdar AP. Curcumin enhances the effects of 5-fluorouracil and oxaliplatin in mediating growth inhibition of colon cancer cells by modulating EGFR and IGF-1R. *Int J Cancer* 2008; 122: 267-273
- 77 Howells LM, Mitra A, Manson MM. Comparison of oxaliplatin- and curcumin-mediated antiproliferative effects in colorectal cell lines. *Int J Cancer* 2007; **121**: 175-183
- 78 Binion DG, Otterson MF, Rafiee P. Curcumin inhibits VEGFmediated angiogenesis in human intestinal microvascular endothelial cells through COX-2 and MAPK inhibition. *Gut* 2008; 57: 1509-1517
- 79 Kunnumakkara AB, Diagaradjane P, Guha S, Deorukhkar A, Shentu S, Aggarwal BB, Krishnan S. Curcumin sensitizes human colorectal cancer xenografts in nude mice to gammaradiation by targeting nuclear factor-kappaB-regulated gene products. *Clin Cancer Res* 2008; 14: 2128-2136
- 80 Rafiee P, Binion DG, Wellner M, Behmaram B, Floer M, Mitton E, Nie L, Zhang Z, Otterson MF. Modulatory effect of curcumin on survival of irradiated human intestinal microvascular endothelial cells: role of Akt/mTOR and NF-{kappa}B. Am J Physiol Gastrointest Liver Physiol 2010; 298: G865-G877
- 81 **Sandur SK**, Deorukhkar A, Pandey MK, Pabón AM, Shentu S, Guha S, Aggarwal BB, Krishnan S. Curcumin modulates the radiosensitivity of colorectal cancer cells by suppressing constitutive and inducible NF-kappaB activity. *Int J Radiat Oncol Biol Phys* 2009; **75**: 534-542
- 82 Su CC, Chen GW, Lin JG, Wu LT, Chung JG. Curcumin inhibits cell migration of human colon cancer colo 205 cells through the inhibition of nuclear factor kappa B / p65 and down-regulates cyclooxygenase-2 and matrix metalloproteinase-2 expressions. *Anticancer Res* 2006; 26: 1281-1288
- 83 Chen A, Xu J. Activation of PPAR{gamma} by curcumin inhibits Moser cell growth and mediates suppression of gene expression of cyclin D1 and EGFR. *Am J Physiol Gastrointest Liver Physiol* 2005; 288: G447-G456
- 84 **Chen A**, Xu J, Johnson AC. Curcumin inhibits human colon cancer cell growth by suppressing gene expression of epi-

dermal growth factor receptor through reducing the activity of the transcription factor Egr-1. *Oncogene* 2006; **25**: 278-287

- 85 Patel BB, Gupta D, Elliott AA, Sengupta V, Yu Y, Majumdar AP. Curcumin targets FOLFOX-surviving colon cancer cells via inhibition of EGFRs and IGF-1R. *Anticancer Res* 2010; 30: 319-325
- 86 Johnson SM, Gulhati P, Arrieta I, Wang X, Uchida T, Gao T, Evers BM. Curcumin inhibits proliferation of colorectal carcinoma by modulating Akt/mTOR signaling. *Anticancer Res* 2009; 29: 3185-3190
- 87 Jaiswal AS, Marlow BP, Gupta N, Narayan S. Beta-cateninmediated transactivation and cell-cell adhesion pathways are important in curcumin (diferuylmethane)-induced growth arrest and apoptosis in colon cancer cells. *Oncogene* 2002; **21**: 8414-8427
- 88 Narayan S. Curcumin, a multi-functional chemopreventive agent, blocks growth of colon cancer cells by targeting betacatenin-mediated transactivation and cell-cell adhesion pathways. J Mol Histol 2004; 35: 301-307
- 89 Ryu MJ, Cho M, Song JY, Yun YS, Choi IW, Kim DE, Park BS, Oh S. Natural derivatives of curcumin attenuate the Wnt/beta-catenin pathway through down-regulation of the transcriptional coactivator p300. *Biochem Biophys Res Commun* 2008; 377: 1304-1308
- 90 Collett GP, Campbell FC. Curcumin induces c-jun N-terminal kinase-dependent apoptosis in HCT116 human colon cancer cells. *Carcinogenesis* 2004; 25: 2183-2189
- 91 Shim JS, Lee J, Park HJ, Park SJ, Kwon HJ. A new curcumin derivative, HBC, interferes with the cell cycle progression of colon cancer cells via antagonization of the Ca2+/calmodulin function. *Chem Biol* 2004; **11**: 1455-1463
- 92 Wang X, Wang Q, Ives KL, Evers BM. Curcumin inhibits neurotensin-mediated interleukin-8 production and migration of HCT116 human colon cancer cells. *Clin Cancer Res* 2006; 12: 5346-5355
- 93 Cruz-Correa M, Shoskes DA, Sanchez P, Zhao R, Hylind LM, Wexner SD, Giardiello FM. Combination treatment with curcumin and quercetin of adenomas in familial adenomatous polyposis. *Clin Gastroenterol Hepatol* 2006; 4: 1035-1038
- 94 Perkins S, Verschoyle RD, Hill K, Parveen I, Threadgill MD, Sharma RA, Williams ML, Steward WP, Gescher AJ. Chemopreventive efficacy and pharmacokinetics of curcumin in the min/+ mouse, a model of familial adenomatous polyposis. *Cancer Epidemiol Biomarkers Prev* 2002; **11**: 535-540
- 95 Tunstall RG, Sharma RA, Perkins S, Sale S, Singh R, Farmer PB, Steward WP, Gescher AJ. Cyclooxygenase-2 expression and oxidative DNA adducts in murine intestinal adenomas: modification by dietary curcumin and implications for clinical trials. *Eur J Cancer* 2006; **42**: 415-421
- 96 Bisht S, Mizuma M, Feldmann G, Ottenhof NA, Hong SM, Pramanik D, Chenna V, Karikari C, Sharma R, Goggins MG, Rudek MA, Ravi R, Maitra A, Maitra A. Systemic administration of polymeric nanoparticle-encapsulated curcumin (NanoCurc) blocks tumor growth and metastases in preclinical models of pancreatic cancer. *Mol Cancer Ther* 2010; 9: 2255-2264
- 97 Swamy MV, Citineni B, Patlolla JM, Mohammed A, Zhang Y, Rao CV. Prevention and treatment of pancreatic cancer by curcumin in combination with omega-3 fatty acids. *Nutr Cancer* 2008; 60 Suppl 1: 81-89
- 98 Ali S, Ahmad A, Banerjee S, Padhye S, Dominiak K, Schaffert JM, Wang Z, Philip PA, Sarkar FH. Gemcitabine sensitivity can be induced in pancreatic cancer cells through modulation of miR-200 and miR-21 expression by curcumin or its analogue CDF. *Cancer Res* 2010; **70**: 3606-3017
- 99 Holcomb B, Yip-Schneider MT, Matos JM, Dixon J, Kennard J, Mahomed J, Shanmugam R, Sebolt-Leopold J, Schmidt CM. Pancreatic cancer cell genetics and signaling response to treatment correlate with efficacy of gemcitabine-based molecular targeting strategies. J Gastrointest Surg 2008; 12: 288-296

- 100 Kunnumakkara AB, Guha S, Krishnan S, Diagaradjane P, Gelovani J, Aggarwal BB. Curcumin potentiates antitumor activity of gemcitabine in an orthotopic model of pancreatic cancer through suppression of proliferation, angiogenesis, and inhibition of nuclear factor-kappaB-regulated gene products. *Cancer Res* 2007; 67: 3853-3861
- 101 Lev-Ari S, Vexler A, Starr A, Ashkenazy-Voghera M, Greif J, Aderka D, Ben-Yosef R. Curcumin augments gemcitabine cytotoxic effect on pancreatic adenocarcinoma cell lines. *Cancer Invest* 2007; 25: 411-418
- 102 Ramachandran C, Resek AP, Escalon E, Aviram A, Melnick SJ. Potentiation of gemcitabine by Turmeric Force in pancreatic cancer cell lines. *Oncol Rep* 2010; 23: 1529-1535
- 103 Lev-Ari S, Zinger H, Kazanov D, Yona D, Ben-Yosef R, Starr A, Figer A, Arber N. Curcumin synergistically potentiates the growth inhibitory and pro-apoptotic effects of celecoxib in pancreatic adenocarcinoma cells. *Biomed Pharmacother* 2005; 59 Suppl 2: S276-S280
- 104 Li L, Braiteh FS, Kurzrock R. Liposome-encapsulated curcumin: in vitro and in vivo effects on proliferation, apoptosis, signaling, and angiogenesis. *Cancer* 2005; 104: 1322-1331
- 105 Mach CM, Mathew L, Mosley SA, Kurzrock R, Smith JA. Determination of minimum effective dose and optimal dosing schedule for liposomal curcumin in a xenograft human pancreatic cancer model. *Anticancer Res* 2009; **29**: 1895-1899
- 106 Mohanty C, Acharya S, Mohanty AK, Dilnawaz F, Sahoo SK. Curcumin-encapsulated MePEG/PCL diblock copolymeric micelles: a novel controlled delivery vehicle for cancer therapy. *Nanomedicine (Lond)* 2010; 5: 433-449
- 107 Li J, Wang Y, Yang C, Wang P, Oelschlager DK, Zheng Y, Tian DA, Grizzle WE, Buchsbaum DJ, Wan M. Polyethylene glycosylated curcumin conjugate inhibits pancreatic cancer cell growth through inactivation of Jab1. *Mol Pharmacol* 2009; 76: 81-90
- 108 Dhillon N, Aggarwal BB, Newman RA, Wolff RA, Kunnumakkara AB, Abbruzzese JL, Ng CS, Badmaev V, Kurzrock R. Phase II trial of curcumin in patients with advanced pancreatic cancer. *Clin Cancer Res* 2008; 14: 4491-4499
- 109 Jutooru I, Chadalapaka G, Lei P, Safe S. Inhibition of NFkappaB and pancreatic cancer cell and tumor growth by curcumin is dependent on specificity protein down-regulation. J Biol Chem 2010; 285: 25332-25344
- 110 Khanbolooki S, Nawrocki ST, Arumugam T, Andtbacka R, Pino MS, Kurzrock R, Logsdon CD, Abbruzzese JL, Mc-Conkey DJ. Nuclear factor-kappaB maintains TRAIL resistance in human pancreatic cancer cells. *Mol Cancer Ther* 2006; 5: 2251-2260
- 111 Li L, Aggarwal BB, Shishodia S, Abbruzzese J, Kurzrock R. Nuclear factor-kappaB and IkappaB kinase are constitutively active in human pancreatic cells, and their down-regulation by curcumin (diferuloylmethane) is associated with the suppression of proliferation and the induction of apoptosis. *Cancer* 2004; **101**: 2351-2362
- 112 **Sun M**, Estrov Z, Ji Y, Coombes KR, Harris DH, Kurzrock R. Curcumin (diferuloylmethane) alters the expression profiles of microRNAs in human pancreatic cancer cells. *Mol Cancer Ther* 2008; **7**: 464-473
- 113 Lev-Ari S, Starr A, Vexler A, Karaush V, Loew V, Greif J, Fenig E, Aderka D, Ben-Yosef R. Inhibition of pancreatic and lung adenocarcinoma cell survival by curcumin is associated with increased apoptosis, down-regulation of COX-2 and EGFR and inhibition of Erk1/2 activity. *Anticancer Res* 2006; 26: 4423-4430
- 114 **Wang Z**, Zhang Y, Banerjee S, Li Y, Sarkar FH. Notch-1 down-regulation by curcumin is associated with the inhibition of cell growth and the induction of apoptosis in pancreatic cancer cells. *Cancer* 2006; **106**: 2503-2513
- 115 Hidaka H, Ishiko T, Furuhashi T, Kamohara H, Suzuki S, Miyazaki M, Ikeda O, Mita S, Setoguchi T, Ogawa M. Curcumin inhibits interleukin 8 production and enhances

interleukin 8 receptor expression on the cell surface:impact on human pancreatic carcinoma cell growth by autocrine regulation. *Cancer* 2002; **95**: 1206-1214

- 116 Glienke W, Maute L, Wicht J, Bergmann L. Curcumin inhibits constitutive STAT3 phosphorylation in human pancreatic cancer cell lines and downregulation of survivin/BIRC5 gene expression. *Cancer Invest* 2010; 28: 166-171
- 117 **Sahu RP**, Batra S, Srivastava SK. Activation of ATM/Chk1 by curcumin causes cell cycle arrest and apoptosis in human pancreatic cancer cells. *Br J Cancer* 2009; **100**: 1425-1433
- 118 Schwer CI, Guerrero AM, Humar M, Roesslein M, Goebel U, Stoll P, Geiger KK, Pannen BH, Hoetzel A, Schmidt R. Heme oxygenase-1 inhibits the proliferation of pancreatic stellate cells by repression of the extracellular signal-regulated kinase1/2 pathway. *J Pharmacol Exp Ther* 2008; **327**: 863-871
- 119 **Glienke W**, Maute L, Wicht J, Bergmann L. Wilms' tumour gene 1 (WT1) as a target in curcumin treatment of pancreatic cancer cells. *Eur J Cancer* 2009; **45**: 874-880
- 120 Hutzen B, Friedman L, Sobo M, Lin L, Cen L, De Angelis S, Yamakoshi H, Shibata H, Iwabuchi Y, Lin J. Curcumin analogue GO-Y030 inhibits STAT3 activity and cell growth in breast and pancreatic carcinomas. *Int J Oncol* 2009; **35**: 867-872
- 121 Friedman L, Lin L, Ball S, Bekaii-Saab T, Fuchs J, Li PK, Li C, Lin J. Curcumin analogues exhibit enhanced growth suppressive activity in human pancreatic cancer cells. *Anticancer Drugs* 2009; **20**: 444-449
- 122 Padhye S, Banerjee S, Chavan D, Pandye S, Swamy KV, Ali S, Li J, Dou QP, Sarkar FH. Fluorocurcumins as cyclooxygenase-2 inhibitor: molecular docking, pharmacokinetics and tissue distribution in mice. *Pharm Res* 2009; 26: 2438-2445
- 123 Xiao J, Chu Y, Hu K, Wan J, Huang Y, Jiang C, Liang G, Li X. Synthesis and biological analysis of a new curcumin analogue for enhanced anti-tumor activity in HepG 2 cells. *Oncol Rep* 2010; 23: 1435-1441
- 124 Wang WZ, Cheng J, Luo J, Zhuang SM. Abrogation of G2/ M arrest sensitizes curcumin-resistant hepatoma cells to apoptosis. FEBS Lett 2008; 582: 2689-2695
- 125 **Cao J**, Jia L, Zhou HM, Liu Y, Zhong LF. Mitochondrial and nuclear DNA damage induced by curcumin in human hepatoma G2 cells. *Toxicol Sci* 2006; **91**: 476-483
- 126 **Cao J**, Liu Y, Jia L, Zhou HM, Kong Y, Yang G, Jiang LP, Li QJ, Zhong LF. Curcumin induces apoptosis through mitochondrial hyperpolarization and mtDNA damage in human hepatoma G2 cells. *Free Radic Biol Med* 2007; **43**: 968-975
- 127 **Jiang MC**, Yang-Yen HF, Lin JK, Yen JJ. Differential regulation of p53, c-Myc, Bcl-2 and Bax protein expression during apoptosis induced by widely divergent stimuli in human hepatoblastoma cells. *Oncogene* 1996; **13**: 609-616
- 128 Bae MK, Kim SH, Jeong JW, Lee YM, Kim HS, Kim SR, Yun I, Bae SK, Kim KW. Curcumin inhibits hypoxia-induced angiogenesis via down-regulation of HIF-1. Oncol Rep 2006; 15: 1557-1562
- 129 Jia L, Wang H, Qu S, Miao X, Zhang J. CD147 regulates vascular endothelial growth factor-A expression, tumorigenicity, and chemosensitivity to curcumin in hepatocellular carcinoma. *IUBMB Life* 2008; **60**: 57-63
- 130 Seol DW, Chen Q, Zarnegar R. Transcriptional activation of the hepatocyte growth factor receptor (c-met) gene by its ligand (hepatocyte growth factor) is mediated through AP-1. *Oncogene* 2000; 19: 1132-1137
- 131 Lin LI, Ke YF, Ko YC, Lin JK. Curcumin inhibits SK-Hep-1 hepatocellular carcinoma cell invasion in vitro and suppresses matrix metalloproteinase-9 secretion. *Oncology* 1998; 55: 349-353
- 132 Notarbartolo M, Poma P, Perri D, Dusonchet L, Cervello M, D'Alessandro N. Antitumor effects of curcumin, alone or in combination with cisplatin or doxorubicin, on human hepatic cancer cells. Analysis of their possible relationship to changes in NF-kB activation levels and in IAP gene expression. *Cancer Lett* 2005; 224: 53-65



- 133 Huang AC, Lin SY, Su CC, Lin SS, Ho CC, Hsia TC, Chiu TH, Yu CS, Ip SW, Lin TP, Chung JG. Effects of curcumin on N-bis(2-hydroxypropyl) nitrosamine (DHPN)-induced lung and liver tumorigenesis in BALB/c mice in vivo. *In Vivo* 2008; 22: 781-785
- 134 Chuang SE, Kuo ML, Hsu CH, Chen CR, Lin JK, Lai GM, Hsieh CY, Cheng AL. Curcumin-containing diet inhibits diethylnitrosamine-induced murine hepatocarcinogenesis. *Carcinogenesis* 2000; 21: 331-335
- 135 Yoysungnoen P, Wirachwong P, Bhattarakosol P, Niimi H, Patumraj S. Antiangiogenic activity of curcumin in hepatocellular carcinoma cells implanted nude mice. *Clin Hemorheol Microcirc* 2005; 33: 127-135
- 136 Yoysungnoen P, Wirachwong P, Bhattarakosol P, Niimi H, Patumraj S. Effects of curcumin on tumor angiogenesis and biomarkers, COX-2 and VEGF, in hepatocellular carcinoma cell-implanted nude mice. *Clin Hemorheol Microcirc* 2006; 34: 109-115
- 137 Yoysungnoen P, Wirachwong P, Changtam C, Suksamrarn A, Patumraj S. Anti-cancer and anti-angiogenic effects of curcumin and tetrahydrocurcumin on implanted hepatocellular carcinoma in nude mice. World J Gastroenterol 2008; 14: 2003-2009
- 138 **Ohashi Y**, Tsuchiya Y, Koizumi K, Sakurai H, Saiki I. Prevention of intrahepatic metastasis by curcumin in an orthotopic implantation model. *Oncology* 2003; **65**: 250-258
- 139 Piper JT, Singhal SS, Salameh MS, Torman RT, Awasthi YC, Awasthi S. Mechanisms of anticarcinogenic properties of curcumin: the effect of curcumin on glutathione linked detoxification enzymes in rat liver. *Int J Biochem Cell Biol* 1998; 30: 445-456
- 140 Sreepriya M, Bali G. Chemopreventive effects of embelin and curcumin against N-nitrosodiethylamine/phenobarbitalinduced hepatocarcinogenesis in Wistar rats. *Fitoterapia* 2005; 76: 549-555
- 141 Sreepriya M, Bali G. Effects of administration of Embelin and Curcumin on lipid peroxidation, hepatic glutathione antioxidant defense and hematopoietic system during N-nitrosodiethylamine/Phenobarbital-induced hepatocarcinogenesis in Wistar rats. *Mol Cell Biochem* 2006; 284: 49-55
- 142 Tang Y, Chen A. Curcumin protects hepatic stellate cells against leptin-induced activation in vitro by accumulating intracellular lipids. *Endocrinology* 2010; 151: 4168-4177
- 143 **Tang Y**, Zheng S, Chen A. Curcumin eliminates leptin's effects on hepatic stellate cell activation via interrupting leptin signaling. *Endocrinology* 2009; **150**: 3011-3020
- 144 Lin J, Zheng S, Chen A. Curcumin attenuates the effects of insulin on stimulating hepatic stellate cell activation by interrupting insulin signaling and attenuating oxidative stress. *Lab Invest* 2009; 89: 1397-1409
- 145 **Zheng S**, Yumei F, Chen A. De novo synthesis of glutathione is a prerequisite for curcumin to inhibit hepatic stellate cell (HSC) activation. *Free Radic Biol Med* 2007; **43**: 444-453
- 146 **Kang Q**, Chen A. Curcumin inhibits srebp-2 expression in activated hepatic stellate cells in vitro by reducing the activity of specificity protein-1. *Endocrinology* 2009; **150**: 5384-5394
- 147 Kang Q, Chen A. Curcumin eliminates oxidized LDL roles in activating hepatic stellate cells by suppressing gene expression of lectin-like oxidized LDL receptor-1. *Lab Invest* 2009; 89: 1275-1290
- 148 Kang Q, Chen A. Curcumin suppresses expression of lowdensity lipoprotein (LDL) receptor, leading to the inhibition of LDL-induced activation of hepatic stellate cells. *Br J Pharmacol* 2009; **157**: 1354-1367
- 149 Lin YL, Lin CY, Chi CW, Huang YT. Study on antifibrotic effects of curcumin in rat hepatic stellate cells. *Phytother Res* 2009; 23: 927-932
- 150 Xu J, Fu Y, Chen A. Activation of peroxisome proliferatoractivated receptor-gamma contributes to the inhibitory effects of curcumin on rat hepatic stellate cell growth. *Am J Physiol Gastrointest Liver Physiol* 2003; 285: G20-G30

- 151 Zheng S, Chen A. Activation of PPARgamma is required for curcumin to induce apoptosis and to inhibit the expression of extracellular matrix genes in hepatic stellate cells in vitro. *Biochem J* 2004; 384: 149-157
- 152 Priya S, Sudhakaran PR. Curcumin-induced recovery from hepatic injury involves induction of apoptosis of activated hepatic stellate cells. *Indian J Biochem Biophys* 2008; 45: 317-325
- 153 Chen A, Zheng S. Curcumin inhibits connective tissue growth factor gene expression in activated hepatic stellate cells in vitro by blocking NF-kappaB and ERK signalling. Br J Pharmacol 2008; 153: 557-567
- 154 Pinlaor S, Prakobwong S, Hiraku Y, Pinlaor P, Laothong U, Yongvanit P. Reduction of periductal fibrosis in liver flukeinfected hamsters after long-term curcumin treatment. *Eur J Pharmacol* 2010; 638: 134-141
- 155 Baghdasaryan A, Claudel T, Kosters A, Gumhold J, Silbert D, Thüringer A, Leski K, Fickert P, Karpen SJ, Trauner M. Curcumin improves sclerosing cholangitis in Mdr2-/- mice by inhibition of cholangiocyte inflammatory response and portal myofibroblast proliferation. *Gut* 2010; **59**: 521-530
- 156 Wu SJ, Tam KW, Tsai YH, Chang CC, Chao JC. Curcumin and saikosaponin a inhibit chemical-induced liver inflammation and fibrosis in rats. *Am J Chin Med* 2010; 38: 99-111
- 157 Fu Y, Zheng S, Lin J, Ryerse J, Chen A. Curcumin protects the rat liver from CCl4-caused injury and fibrogenesis by attenuating oxidative stress and suppressing inflammation. *Mol Pharmacol* 2008; **73**: 399-409
- 158 Wu SJ, Lin YH, Chu CC, Tsai YH, Chao JC. Curcumin or saikosaponin a improves hepatic antioxidant capacity and protects against CCl4-induced liver injury in rats. J Med Food 2008; 11: 224-229
- 159 Shu JC, He YJ, Lv X, Ye GR, Wang LX. Curcumin prevents liver fibrosis by inducing apoptosis and suppressing activation of hepatic stellate cells. J Nat Med 2009; 63: 415-420
- 160 Bruck R, Ashkenazi M, Weiss S, Goldiner I, Shapiro H, Aeed H, Genina O, Helpern Z, Pines M. Prevention of liver cirrhosis in rats by curcumin. *Liver Int* 2007; 27: 373-383
- 161 Vizzutti F, Provenzano A, Galastri S, Milani S, Delogu W, Novo E, Caligiuri A, Zamara E, Arena U, Laffi G, Parola M, Pinzani M, Marra F. Curcumin limits the fibrogenic evolution of experimental steatohepatitis. *Lab Invest* 2010; **90**: 104-115
- 162 Hanai H, Iida T, Takeuchi K, Watanabe F, Maruyama Y, Andoh A, Tsujikawa T, Fujiyama Y, Mitsuyama K, Sata M, Yamada M, Iwaoka Y, Kanke K, Hiraishi H, Hirayama K, Arai H, Yoshii S, Uchijima M, Nagata T, Koide Y. Curcumin maintenance therapy for ulcerative colitis: randomized, multicenter, double-blind, placebo-controlled trial. *Clin Gastroenterol Hepatol* 2006; **4**: 1502-1506
- 163 Holt PR, Katz S, Kirshoff R. Curcumin therapy in inflammatory bowel disease: a pilot study. *Dig Dis Sci* 2005; 50: 2191-2193
- 164 Epstein J, Docena G, MacDonald TT, Sanderson IR. Curcumin suppresses p38 mitogen-activated protein kinase activation, reduces IL-1beta and matrix metalloproteinase-3 and enhances IL-10 in the mucosa of children and adults with inflammatory bowel disease. Br J Nutr 2010; 103: 824-832
- 165 Larmonier CB, Uno JK, Lee KM, Karrasch T, Laubitz D, Thurston R, Midura-Kiela MT, Ghishan FK, Sartor RB, Jobin C, Kiela PR. Limited effects of dietary curcumin on Th-1 driven colitis in IL-10 deficient mice suggest an IL-10-dependent mechanism of protection. *Am J Physiol Gastrointest Liver Physiol* 2008; 295: G1079-G1091
- 166 Ung VY, Foshaug RR, MacFarlane SM, Churchill TA, Doyle JS, Sydora BC, Fedorak RN. Oral administration of curcumin emulsified in carboxymethyl cellulose has a potent antiinflammatory effect in the IL-10 gene-deficient mouse model of IBD. *Dig Dis Sci* 2010; 55: 1272-1277
- 167 **Nones K**, Dommels YE, Martell S, Butts C, McNabb WC, Park ZA, Zhu S, Hedderley D, Barnett MP, Roy NC. The effects of dietary curcumin and rutin on colonic inflammation

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and gene expression in multidrug resistance gene-deficient (mdr1a-/-) mice, a model of inflammatory bowel diseases. *Br J Nutr* 2009; **101**: 169-181

- 168 Salh B, Assi K, Templeman V, Parhar K, Owen D, Gomez-Munoz A, Jacobson K. Curcumin attenuates DNB-induced murine colitis. Am J Physiol Gastrointest Liver Physiol 2003; 285: G235-G243
- 169 Martelli L, Ragazzi E, di Mario F, Martelli M, Castagliuolo I, Dal Maschio M, Palu G, Maschietto M, Scorzeto M, Vassanelli S, Brun P. A potential role for the vanilloid receptor TRPV1 in the therapeutic effect of curcumin in dinitrobenzene sulphonic acid-induced colitis in mice. *Neurogastroenterol Motil* 2007; 19: 668-674
- 170 Lubbad A, Oriowo MA, Khan I. Curcumin attenuates inflammation through inhibition of TLR-4 receptor in experimental colitis. *Mol Cell Biochem* 2009; **322**: 127-135
- 171 **Lubbad AS**, Oriowo MA, Khan I. Curcumin reverses attenuated carbachol-induced contraction of the colon in a rat model of colitis. *Scand J Gastroenterol* 2009; **44**: 187-194
- 172 Jian YT, Mai GF, Wang JD, Zhang YL, Luo RC, Fang YX. Preventive and therapeutic effects of NF-kappaB inhibitor curcumin in rats colitis induced by trinitrobenzene sulfonic acid. World J Gastroenterol 2005; 11: 1747-1752
- 173 Camacho-Barquero L, Villegas I, Sánchez-Calvo JM, Talero E, Sánchez-Fidalgo S, Motilva V, Alarcón de la Lastra C. Curcumin, a Curcuma longa constituent, acts on MAPK p38 pathway modulating COX-2 and iNOS expression in chronic experimental colitis. *Int Immunopharmacol* 2007; 7: 333-342
- 174 Jiang H, Deng CS, Zhang M, Xia J. Curcumin-attenuated trinitrobenzene sulphonic acid induces chronic colitis by inhibiting expression of cyclooxygenase-2. World J Gastroenterol 2006; 12: 3848-3853
- 175 Billerey-Larmonier C, Uno JK, Larmonier N, Midura AJ, Timmermann B, Ghishan FK, Kiela PR. Protective effects of dietary curcumin in mouse model of chemically induced colitis are strain dependent. *Inflamm Bowel Dis* 2008; 14: 780-793
- 176 Venkataranganna MV, Rafiq M, Gopumadhavan S, Peer G, Babu UV, Mitra SK. NCB-02 (standardized Curcumin preparation) protects dinitrochlorobenzene- induced colitis through down-regulation of NFkappa-B and iNOS. World J Gastroenterol 2007; 13: 1103-1107
- 177 Arafa HM, Hemeida RA, El-Bahrawy AI, Hamada FM. Pro-

phylactic role of curcumin in dextran sulfate sodium (DSS)induced ulcerative colitis murine model. *Food Chem Toxicol* 2009; **47**: 1311-1317

- 178 Yadav VR, Suresh S, Devi K, Yadav S. Effect of cyclodextrin complexation of curcumin on its solubility and antiangiogenic and anti-inflammatory activity in rat colitis model. *AAPS PharmSciTech* 2009; **10**: 752-762
- 179 Yadav VR, Suresh S, Devi K, Yadav S. Novel formulation of solid lipid microparticles of curcumin for anti-angiogenic and anti-inflammatory activity for optimization of therapy of inflammatory bowel disease. J Pharm Pharmacol 2009; 61: 311-321
- 180 Larmonier CB, Midura-Kiela MT, Ramalingam R, Laubitz D, Janikashvili N, Larmonier N, Ghishan FK, Kiela PR. Modulation of neutrophil motility by curcumin: Implications for inflammatory bowel disease. *Inflamm Bowel Dis* 2010; 17: 503-515
- 181 Binion DG, Heidemann J, Li MS, Nelson VM, Otterson MF, Rafiee P. Vascular cell adhesion molecule-1 expression in human intestinal microvascular endothelial cells is regulated by PI 3-kinase/Akt/MAPK/NF-kappaB: inhibitory role of curcumin. Am J Physiol Gastrointest Liver Physiol 2009; 297: G259-G268
- 182 Ukil A, Maity S, Karmakar S, Datta N, Vedasiromoni JR, Das PK. Curcumin, the major component of food flavour turmeric, reduces mucosal injury in trinitrobenzene sulphonic acid-induced colitis. Br J Pharmacol 2003; 139: 209-218
- 183 Sugimoto K, Hanai H, Tozawa K, Aoshi T, Uchijima M, Nagata T, Koide Y. Curcumin prevents and ameliorates trinitrobenzene sulfonic acid-induced colitis in mice. *Gastroenterology* 2002; 123: 1912-1922
- 184 Shoba G, Joy D, Joseph T, Majeed M, Rajendran R, Srinivas PS. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med* 1998; 64: 353-356
- 185 Bisht S, Feldmann G, Soni S, Ravi R, Karikar C, Maitra A, Maitra A. Polymeric nanoparticle-encapsulated curcumin ("nanocurcumin"): a novel strategy for human cancer therapy. J Nanobiotechnology 2007; 5: 3
- 186 Sahu A, Kasoju N, Goswami P, Bora U. Encapsulation of Curcumin in Pluronic Block Copolymer Micelles for Drug Delivery Applications. J Biomater Appl 2010; Epub ahead of print

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